Original article

Differential ligand-signaling network of CCL19/CCL21-CCR7 system

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Abstract

Chemokine (C-C motif) receptor 7 (CCR7), a class A subtype G-Protein Coupled Receptor (GPCR), is involved in the migration, activation and survival of multiple cell types including dendritic cells, T cells, eosinophils, B cells, endothelial cells and different cancer cells. Together, CCR7 signaling system has been implicated in diverse biological processes such as lymph node homeostasis, T cell activation, immune tolerance, inflammatory response and cancer metastasis. CCL19 and CCL21, the two well-characterized CCR7 ligands, have been established to be differential in their signaling through CCR7 in multiple cell types. Although the differential ligand signaling through single receptor have been suggested for many receptors including GPCRs, there exists no resource or platform to analyse them globally. Here, first of its kind, we present the cell-type-specific differential signaling network of CCL19/CCL21-CCR7 system for effective visualization and differential analysis of chemokine/GPCR signaling.

Database URL: http://www.netpath.org/pathways?path_id=NetPath_46.

Introduction

CCL19 and CCL21, also known as EBI1-Ligand Chemokine/Macrophage Inflammatory Protein-3β (ELC/MIP-3β) and Secondary Lymphoid-tissue Chemokine (SLC), respectively, are the major chemokines predominantly expressed in secondary lymphoid tissues (1–3). Elevated expression of the chemokines CCL19 and CCL21 have been reported in diverse disease conditions such as atherosclerosis (4, 5), cancers, bone disorders (6, 7) and other inflammatory conditions such as asthma (8), HIV infection (9) and pneumonia (10). Structurally, CCL21 differ from CCL19 in that it has an extra 32 amino acid C-terminus of basic amino acids that may mediate the distinct binding of CCL21 to other molecules (11, 12). Although,

CCL19 and/or CCL21 have also been reported to bind to cell surface receptors such as chemokine (C-C motif) receptor-like 2 (CCRL2) and atypical chemokine receptor 4 (ACKR4), their functional effects are identified to be mediated through the class-A subtype GPCR, Chemokine (C-C motif) receptor 7 (CCR7) (13–16). Facilitating the migration, activation and survival of multiple cell types including dendritic cells, T cells, eosinophils, B cells, endothelial cells and different cancer cells, CCR7 signaling system has been implicated in diverse biological processes such as lymph node homeostasis, T cell activation, immune tolerance, inflammatory response and cancer metastasis (17–19). Dendritic cell- and stromal cell-based intratumoral delivery of CCL21 for enhanced recruitment and activation of antigen presenting cells and T cells are being evaluated for lung cancer therapy (20, 21).

Zidar *et al*. (22) have reported the differential and selective activation of the members of the G-protein-coupled receptor kinase (GRK)/beta-arrestin system of CCR7 by its two ligands, CCL19 and CCL21, with no substantial bias in the inhibition of adenylate cyclase by the CCR7-$G\alpha_{i/o}$ system in HEK293 cells. Recently, Corbisier *et al*. (23) have reported based on bioluminescence resonance energy transfer assays that there is a ligand bias in the activation of multiple $G\alpha_{i/o}$ isoforms of CCR7 in HEK293 cells. Subsequently, a large number of molecules and their involvement in mediating diverse biological processes of CCR7 have been analysed and reported under stimulation with either or both of these ligands in multiple cell types. With no further ligands currently established for CCR7 and taking into account the differences in sequences and their differential existence in soluble form as discussed by Comerford *et al*. (24), it is intriguingly significant to analyse the differential signaling between CCL19 and CCL21 in multiple cell types. To analyse the differential signaling of CCR7 ligands further, the experimentally proven molecular reactions induced by CCR7 ligands should be made available in computationally analysable formats such as BioPAX, SBML or GPML formats in a cell-type-specific manner (25–27).

To this end, we have compiled the first CCR7 signaling pathway and have made available the data in multiple exchangeable and analysable formats. The first of its kind, we have also have analysed and represented the experimentally reported molecular information available in CCR7 signaling to develop the cell-type-specific differential ligand-signaling network of CCR7.

Curation of CCR7 Signaling Reactions

The genes and reactions reported to be induced by CCL19 or CCL21 through CCR7 were characterized into binary and complex protein–protein interactions, direct and indirect enzyme–substrate reactions, activation and inhibition status of proteins, protein translocations across multiple cellular compartments and genes differentially regulated at transcriptional and translational level, each of them following the criteria detailed before (28–30). The direct enzyme–substrate reactions should also be considered as protein–protein interactions. For each of the reactions, the specific cell types in which the reactions are studied were also documented. PathBuilder, an in-house pathway curation tool that facilitates quality check and thereby error-free curation was used to document all the reactions systematically (31).

Identification of Differential-Ligand Induced Reactions

The different types of reactions aforementioned induced by similar dosage of CCL19 and/or CCL21 through CCR7 were analysed to identify reactions that were differentially induced by these ligands. We have currently characterized the differential-ligand induced reactions to a cell-type-specific manner. The reactions that were studied and identified to be induced by any one of the ligands without proof for the other ligand in a particular cell type are not considered as differential.

Table 1. The reactions reported to be differentially regulated by CCL19 or CCL21[a]

CCR7 signaling pathway reactions	CCL19	CCL21	Cell lines/types
CCR7-ADBRK2 recruitment	blue		HEK293 cells
CCR7-Beta arrestin 2 recruitment	red		HEK293 cells
CCR7-Arrestin 3 recruitment	blue		HuT78 cells
Arrestin 2 mediated internalization	red		HEK293 cells
Arrestin 3 mediated internalization	blue		HuT78 cells
GNAI1 activation	red		HEK293 cells
GNAI2 activation	red		HEK293 cells
GNAI3 activation	red		HEK293 cells
GNAO activation	red		HEK293 cells
PIK3R1 activation	red		Mature DCs
PIK3R1 activation		red	Normal B cells
AKT1 activation	red		Mature DCs
AKT1 activation		red	CD8+ T cells
ERK1/2 activation	red		HEK293 cells
ERK5 activation	red		HuT78 cells
Activation of KLF2	blue		HuT78 cells

[a]The reactions differentially regulated by CCL19 or CCL21 in multiple cell types are distinguished by color codes. Red color box indicates that the corresponding ligand induces higher activation of the reaction than the other. Blue color box indicates that the reactions are identified to be induced solely by CCL19 and not by CCL21 in the specific cell lines studied.

Table 2. The proteins reported to be activated by CCL19 or CCL21 in normal and multiple cancer cell types[a]

Proteins Gene symbol	Ligands CCL19/21 19	21	Normal cells Cell type	Proteins Gene symbol	Ligands CCL19/21 19	21	Cancer cells Cell lines	Proteins Gene symbol	Ligands CCL19/21 19	21	HNSCC cells Cell line
MAPK14			B cells	CCNB1			A460 cells	PLCG1			PCI-15B cells
PIK3R1			B cells	CCNA1			A460 cells	AKT1			PCI-15B cells
JAK3			CD4+ T cells	CDK1			A460 cells	PLCG1			PCI-37B cells
JAK3			CD8+ T cells	MAPK1			A460 cells	PRKCA			PCI-37B cells
AKT1			CD8+ T cells	MAPK3			A460 cells	RHOA			PCI-37B cells
BCL2L1			CD8+ T cells	MAPK8			A460 cells	JAK2			PCI-37B cells
PLCG2			Dentritic cells	MAPK9			A460 cells	SRC			PCI-37B cells
ANO6			Dentritic cells	MAPK14			A460 cells	ROCK1			PCI-37B cells
MTOR			Dentritic cells	SP1			A549 cells	PTK2B			PCI-37B cells
RPS6KB1			Dentritic cells	HPSE			A549 cells	CFL1			PCI-37B cells
EIF4EBP1			Dentritic cells	CCNB1			A549 cells	STAT3			PCI-37B cells
RELA			Dentritic cells	CCNA1			A549 cells	AKT1			PCI-37B cells
BCL2L1			Dentritic cells	CDK1			A549 cells	MTOR			PCI-37B cells
MAP2K1			Dentritic cells	MAPK1			A549 cells	RPS6KB1			PCI-37B cells
MAP2K2			Dentritic cells	MAPK3			A549 cells	MMP9			PCI-37B cells
MAPK1			Dentritic cells	MAPK8			A549 cells	VIM			PCI-37B cells
MAPK3			Dentritic cells	MAPK9			A549 cells	MAPK1			PCI-37B cells
MAPK8			Dentritic cells	MAPK14			A549 cells	MAPK3			PCI-37B cells
MAPK9			Dentritic cells	RHOA			B-CLL cells	MAPK8			PCI-37B cells
MAPK14			Dentritic cells	ROCK1			B-CLL cells	MAPK9			PCI-37B cells
RHOA			Mature DCs	MYL2			B-CLL cells	MAPK14			PCI-37B cells
RAC1			Mature DCs	AKT1			B-CLL cells	PLCG1			PCI-4B cells
CDC42			Mature DCs	FOS			B-CLL cells	JAK2			PCI-4B cells
ROCK1			Mature DCs	ATF2			B-CLL cells	SRC			PCI-4B cells
PTK2B			Mature DCs	MMP9			B-CLL cells	PTK2B			PCI-4B cells
CFL1			Mature DCs	MAPK1			B-CLL cells	STAT3			PCI-4B cells
AKT1			Mature DCs	MAPK3			B-CLL cells	AKT1			PCI-4B cells
RELA			Mature DCs	MAPK8			B-CLL cells	MTOR			PCI-4B cells
MAP2K4			Mature DCs	MAPK9			B-CLL cells	RPS6KB1			PCI-4B cells
MAPK8			Mature DCs	MAPK14			B-CLL cells	RELA			PCI-4B cells
MAPK9			Mature DCs	PIK3R1			B-CLL cells	VIM			PCI-4B cells
MAPK14			Mature DCs	GNAI1			HEK293 cells	MAPK1			PCI-4B cells
PIK3R1			Mature DCs	GNAI2			HEK293 cells	MAPK3			PCI-4B cells
AKT1			Mesangial cells	GNAI3			HEK293 cells	MAPK8			PCI-4B cells
PLCG1			Naïve T cells	GNAO			HEK293 cells	MAPK9			PCI-4B cells
JAK3			PBLs	MAPK1			HEK293 cells	MAPK14			PCI-4B cells
PRKCQ			T cells	MAPK3			HEK293 cells				
EZR			T cells	ADBRK1			HEK293 cells				
MMP9			BE1 cells	GRK6			HEK293 cells				
				ARRB2			HEK293 cells				
				KLF2			HuT78 cells				
				SIPR1			HuT78 cells				
				MAPK7			HuT78 cells				
				ARRB2			HuT78 cells				
				ARR3			HuT78 cells				

[a]The proteins regulated by CCL19 or CCL21 in multiple normal cells and cancer cell lines are distinguished by color codes. Red color box indicates that the corresponding ligand induces relatively higher activity of the protein than the other by virtue of increased activating post-translational modified forms with no significant differences in the protein amount. Blue color box indicates that the reactions are identified to be induced solely by CCL19 and not by CCL21 in the specific cell lines studied. The green color box indicates that the activity of the specific molecules is studied in particular cell types only for the corresponding ligand.

Development of a Cell-Type-Specific Differential-Ligand Signaling Network of CCR7

Molecular reactions reported in CCR7 signaling pathway

Manual analysis of the CCR7 signaling pathway reactions from 65 research articles out of a total of over 3000 articles that were screened resulted in the documentation of 24 protein–protein interaction events, 45 enzyme–substrate events, 13 protein translocation events. Together, we identified that 45 molecules were found to be activated and 9 proteins to be inhibited in CCR7 signaling for the dosage and time points reported in the published research articles. We have also documented 19 genes to be upregulated and 3 genes to be downregulated at the transcriptional and/or translational level.

Cell-type-specific differential-ligand specific reactions

The analysis of the CCR7 signaling pathway reactions have resulted in the identification of 14 reactions to be differentially regulated in five cell types. For the other cell types, either CCL19 or CCL21 have been used to study the induced reactions (most of them belonged to this category) or that the differential signaling has not been identified and rather been reported as induced by both the ligands. The reactions that were identified to be differentially regulated are summarized in Table 1. In addition, the proteins reported to be activated in normal cells and multiple cancer cell types are provided in Table 2.

Visualization of the CCR7 signaling pathway

The pathway was manually represented using the visualization tool named 'PathVisio' (27). The topology was deduced based on the information obtained from assays using inhibitors/activators, silencing approaches, mutants and *in vitro* kinase assays. Specific reactions are denoted by edges connecting the nodes as represented in the legend in Figure 1. The differential signaling reactions in specific cell types have been represented using square, triangle and circle shapes distinguished by colors. Apart from the genes and their reactions, the biological processes or functions
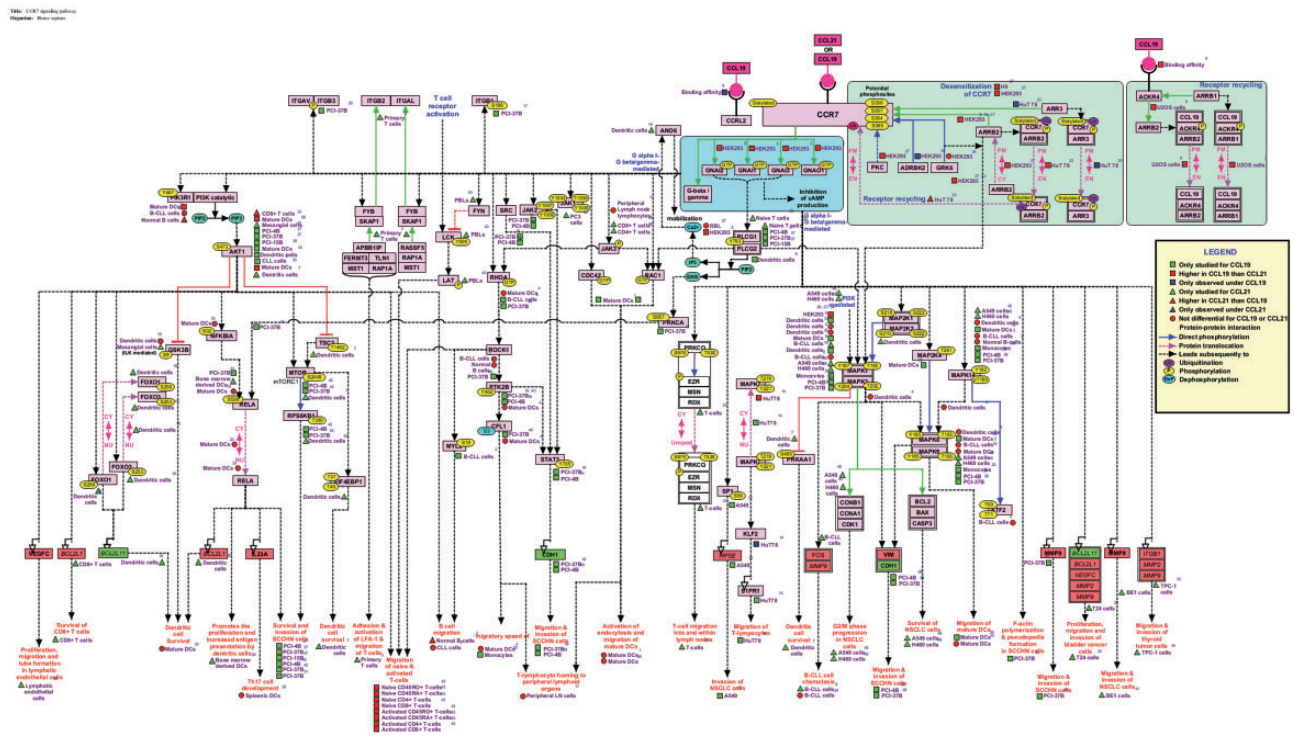

Figure 1. Cell-type-specific differential-ligand network map of CCL19/CCL21-CCR7 signaling. The network map of CCR7 signaling pathway reactions induced by its ligands CCL19 and CCL21 that are experimentally identified in multiple cell types are represented using PathVisio. The topology of the network is devised based on the mutants, inhibitors, activators and silencing approaches. The edges represent the reactions and nodes represent the molecules. Molecules italicized represent the genes transcriptionally regulated by CCR7 signaling with red indicating the upregulated genes and green, the downregulated genes. The edges are distinguished by colors to differentiate different types of reactions such as protein–protein interactions, Post-translational modifications (PTMs) and translocation reactions. The PTMs of each protein are represented with the site and residue of modification mapped to the corresponding RefSeq accessions. The differential signaling by CCL19 and CCL21 in multiple cell types are also represented using colored square (CCL19), triangle (CCL21) and circle (not differential for both CCL19 and CCL21) shapes. Blue represent the reactions to be unique to one of them but not induced by the other whereas red indicates higher for one and the green indicates studied only in one of them and not the other and hence not to be considered differential.

that were assayed to be mediated by the genes were also compiled with their specificity to cell types and differential-ligand stimulation whenever available.

Data formats, availability and visualization

The CCR7 pathway data is free to the scientific community through the resource of signaling pathways, NetPath, at http://www.netpath.org/pathways?path_id=NetPath_46. The description to each molecular reactions and more information on specific molecules are available through the molecule pages in NetPath. The CCR7 signaling pathway data can be downloaded from NetPath in multiple data exchangeable and analysable formats such Proteomics Standards Initiative for Molecular Interaction (PSI-MI version 2.5) (32), Biological PAthway eXchange (BioPAX level 3) (25) and Systems Biology Markup Language (SBML level 2.1) (26). The gpml file can also be downloaded for visualization, analysis and editing from the NetSlim page in NetPath (http://www.netpath.org/netslim/ccr7_pathway.html). GPML format can be used for pathway enrichment analysis using GeneSpring (Agilent). The CCR7 signaling data provided in these formats may also be visualized and analysed using the software such as Cytoscape (33), GenMAPP, Visualization and layout services for BioPAX pathway models (VISIBIOweb) (34) and Chisio BioPAX Editor (ChiBE) (35). The pathway data will be updated periodically as more reactions become available.

Conclusions

Signaling network maps are essential template for pathway analysis of high-throughput data. We have already developed over 40 signaling pathways facilitating this purpose and have made them available through NetPath. Here, we report not only the first chemokine signaling pathway map developed through the NetPath criteria but also the first ligand-specific and cell-type-specific signaling network induced differentially by two ligands of a single receptor, CCR7, the CCL19 and CCL21. We believe that this signaling pathway will serve as an efficient platform for the further analysis of the differential signaling between CCL19/CCL21 and also the reference for development of detailed maps for analysis of many more signaling pathways including GPCRs in the future.

Funding

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