

Database, 2019, 1–7 doi: 10.1093/database/baz027 Database tool



Database tool

# SELER: a database of super-enhancer-associated IncRNA- directed transcriptional regulation in human cancers

Zhi-Wei Guo<sup>1,†</sup>, Chen Xie<sup>2,†</sup>, Kun Li<sup>1</sup>, Xiang-Ming Zhai<sup>1</sup>, Geng-Xi Cai<sup>3</sup>, Xue-Xi Yang<sup>1</sup> and Ying-Song Wu<sup>1,\*</sup>

<sup>1</sup>Institute of Antibody Engineering, School of Laboratory Medicine and Biotechnology, Southern Medical University, Guangzhou 510515, China, <sup>2</sup>Key Laboratory of Liver Disease of Guangdong Province, The Third Affiliated Hospital of Sun Yat-sen University, Guangzhou 510630, China and <sup>3</sup>Department of Breast Surgery, The First People's Hospital of Foshan, Foshan 528000, China

\*Corresponding author: Tel: +86-20-61648550; Fax: +86-20-61648554; Email: wuyingsong2017@126.com <sup>†</sup>Zhi-Wei Guo and Chen Xie are co-first authors.

Citation details: Guo,Z.-W., Xie,C., Li,K. *et al.* SELER: a database of super-enhancer-associated lncRNA- directed transcriptional regulation in human cancers. *Database* (2019) Vol. 2019: article ID baz027; doi:10.1093/database/baz027

Received 5 June 2018; Revised 2 February 2019; Accepted 6 February 2019

# Abstract

Super-enhancers (SEs) are enriched with a cluster of mediator binding sites, which are major contributors to cell-type-specific gene expression. Currently, a large quantity of long non-coding RNAs has been found to be transcribed from or to interact with SEs, which constitute super-enhancer associated long non-coding RNAs (SE-IncRNAs). These SE-IncRNAs play essential roles in transcriptional regulation through controlling SEs activity to regulate a broad range of physiological and pathological processes, especially tumorigenesis. However, the pathological functions of SE-IncRNAs in tumorigenesis are still obscure. In this paper, we characterized 5056 SE-IncRNAs and their associated genes by analysing 102 SE data sets. Then, we analysed their expression profiles and prognostic information derived from 19 cancer types to identify cancer-related SE-IncRNAs and to explore their potential functions. In total, 436 significantly differentially expressed SE-IncRNAs and 2035 SE-IncRNAs with high prognostic values were identified. Additionally, 3935 significant correlations between SE-IncRNAs and their regulatory genes were further validated by calculating their correlation coefficients in each cancer type. Finally, the SELER database incorporating the aforementioned data was provided for users to explore their physiological and pathological functions to comprehensively understand the blocks of living systems.

Database URL: www.seler.cn

## Introduction

Super-enhancers (SEs) are enriched with clustered mediator binding sites and a variety of chromatin signatures, such as H3K4me1, H3K4me3, H3K27ac and P300 acetyltransferase, which play essential roles in regulating gene expression (1-3). The enriched chromatin signature could reflect the regulatory roles of genomic regions; therefore, they could be applied to identify SEs (2). SEs exist in a wide range of mammalian cells, and they can increase gene transcription over large genomic distances to regulate gene expression and to determine cell-type specificity (2, 4). More importantly, SEs are closely related with a variety of diseases, especially human cancers (5, 6). For instance, SEs have been shown to affect the invasion and metastasis of neuroendocrine tumor cells by controlling MET expression (7). As SEs play important roles in controlling gene expression to regulate cellular physiological and pathological processes, it is necessary to reveal their underlying regulatory mechanisms.

Currently, pervasive transcriptions of the human genome have been documented, and most of them are non-coding transcripts, especially long non-coding RNAs (lncRNAs), which are endogenous non-coding RNAs that are longer than 200 nucleotides (nt) (8, 9). LncRNAs have been proven to play essential roles in regulating the expression of genes that affect numerous biological processes, such as the cell cycle and apoptosis (10, 11). Recent discoveries have revealed that lncRNAs transcribed from or that are interact with SE regulatory elements constitute a specific type of lncRNAs, which were termed as super-enhancer associated lncRNAs (SE-lncRNAs) (12, 13). SE-lncRNAs regulate gene expression by affecting gene promoter activity (14-16). Although SE-lncRNAs significantly contributed to gene expression, the systematic identification of SElncRNAs and their regulated genes still lacks comprehensive recognition.

SE-lncRNAs have been proven to play essential roles in regulating physiological and pathological processes, especially tumorigenesis. For instance, SE-lncRNA cardiac mesoderm enhancer-associated noncoding RNA (CAR-MEN) is upregulated during the development process and it controls cardiac precursor cell differentiation (15). Moreover, upperhand can regulate heart development by affecting hand2 expression levels (14). In addition to regulating physiological processes, SE-lncRNAs are closely correlated with tumorigenesis (16). SE-lncRNA CCAT1-L promotes cancer growth by forming enhancer loops to activate the MYC expression (16). Despite of their critical roles in various physiological and pathological processes, their potential roles in human cancers still lack comprehensive investigation. To systematically explore the potential regulatory roles of SE-lncRNAs in tumor progression, we developed <u>SE-lncRNA</u> directed transcriptional regulation in the human canc<u>ers</u> (SELER) database. SELER first identified putative SE-lncRNAs and their associated genes. More importantly, their potential functions in cancers were further explored by analysing their expression profiles, correlation coefficient and prognostic value across 19 cancer types. Finally, SELER was built to store and display data.

### Methods

The analytical workflow of the construction of the cancerrelated SE-lncRNA database mainly consisted of the following three sections: SE-lncRNA identification, cancerrelated SE-lncRNA annotation and database construction (Figure 1).

## SE-IncRNA identification

The SEs of 102 different cell lines and the lncRNA information were downloaded from dbSUPER (downloaded on 1 October 2018) (2) and GENCODE (v27) (17), respectively. The human reference genome (hg19) was applied to handle genomic coordinates. By comparing genomic coordinates, the whole length or the transcription start site (TSS) of an lncRNA within an SE was taken as a cis-acting SE-lncRNA. A previous study has identified hundreds of trans-acting SE-lncRNAs, which were downloaded from its Supplementary Materials (13). As trans-acting SE-lncRNAs mainly exerted their functions in the nucleus, we applied lncLocator (18) to predict nuclear-retained lncRNAs. Then, the interactions between SEs and trans-acting SE-lncRNAs were predicted using Triplexator with options: -1 19 -e 5 -c 1 (19). As the previous study revealed, most of regulated genes of SEs were within 50 kilobase (kb) (20), and the TSSs of the protein-coding genes covered by SEs or within a segment 50 kb upstream or downstream of the SEs were taken as their regulated genes. Functional enrichment analysis was performed using PANTHER with the default setting to analyse the regulated genes of two types of SE-lncRNAs (Supplementary Material, Table 1) (21).

## Cancer-related SE-IncRNA annotation

The lncRNA expression profiles, Pearson's correlation coefficient between lncRNA and protein coding genes and prognostic values across 19 cancer types were downloaded from TANRIC (v1.0) (22). By comparing the SE-lncRNAs



**Figure 1.** System overview of cancer-related SE-IncRNAs database construction. The workflow of cancer-related SE-IncRNA database construction mainly consisted of the following three sections: SE-IncRNA identification, cancer-related SE-IncRNA annotation and database construction. We first identified trans-acting and cis-acting SE-IncRNAs according to their regulatory mechanisms (left part of Figure 1). To explore cancer-related SE-IncRNAs, we identified significantly differentially expressed SE-IncRNAs and SE-IncRNAs with high prognostic values (right part of Figure 1). Moreover, we calculated the correlation coefficient along with the regulated genes of each cancer type to identify their truly regulatory relationships. Finally, the SELER database was built.

expression levels in cancer tissues with their corresponding adjacent normal tissues, differentially expressed SElncRNAs were identified by using Student's *t*-test. The *P*-value was adjusted to the false discovery rate (FDR) by using the Benjamini–Hochberg method. FDR  $\leq 0.1$ and llog<sub>2</sub>fold changel  $\geq 1$  were taken as the criteria to identify significantly differentially expressed lncRNAs. We identified SE-lncRNAs with highly significant prognostic value (*P*-value  $\leq 0.05$ ) from the survival analysis results of TANRIC. To annotate the functions of SE-lncRNAs in cancers or other diseases, the databases of LncRNADisease (11) and Lnc2Cancer (10) were used (downloaded on 1 May 2018).

#### Database construction

To store and display data, a LAMP (Linux, Apache, MySQL and PHP)-based database and web server were developed, which were provided for users to explore the potential regulatory functions of cancer-related SE-lncRNAs.

## Results

#### Putative regulatory genes of SE-IncRNAs

Previous studies have reported that two types of SE-lncRNAs exist, including cis-acting and trans-acting SE-lncRNAs (2, 12). The cis-acting SE-lncRNAs were

Mechanism	Super enhancer	lncRNA gene	lncRNA transcript	Gene	Gene transcript	Regulatory relationshi				
Cis-acting	24 697	4996	8821	8171	57 843	16 272				
Trans-acting	4629	123	179	4577	32 676	11 214				
Total	27 029	5056	8908	9491	67 899	27 481				

Table 1. Data statistics of SE-IncRNAs and their regulated genes

Gene = regulated genes of SE-lncRNAs. Gene transcript = regulated gene transcripts of SE-lncRNAs. Regulatory relationship = regulatory relations between SE-lncRNA and their regulated genes.

 Table 2.
 Data statistics of cancer-related SE-IncRNAs

Туре	DF	PV	DF&PV	Significant relationship
Cis-acting	430	2032	347	3622
Trans-acting	13	19	7	401
Total	436	2035	349	3935

DF = significantly differentially expressed SE-lncRNAs. PV = SE-lncRNAs with a high prognostic value. DF&PV = significantly differentially expressed SE-lncRNAs with a high prognostic value. Significant relationship = significant regulatory relationship with *P*-value $\leq$ 0.05 in Cox's proportional hazard analysis.

transcribed from SEs to regulate the gene nearby, and the trans-acting SE-lncRNAs were transcribed from other genomic coordinates that interact with SEs to regulate genes from a distance. In total, we identified 5056 SE-lncRNAs, including 4996 cis-acting and 123 transacting SE-lncRNAs (Table 1). The cis-acting SE-lncRNAs were transcribed from 24 697 SEs and the trans-acting SE-lncRNAs interacted with 4629 SEs to regulate closet gene expression. By comparing the genome coordinates of SEs with the closet protein-coding genes, 8171 and 4557 regulated genes were inferred to be regulated by cisand trans-acting SE-lncRNAs, respectively (Table 1). Then, we investigated the predominantly regulatory functions of the regulated genes of two types of SE-lncRNAs by gene functional enrichment analysis. The enriched results showed that the regulated genes of cis-acting and transacting SE-IncRNAs were enriched in the immune-related processes, indicating their potential roles in regulating human immune system (Supplementary Material, Table 2 and Supplementary Material, Table 3).

# Substantial cancer-related SE-IncRNAs identified in multiple cancers

To recognize cancer-related SE-lncRNAs, we first identified significantly differentially expressed SE-lncRNAs and SE-lncRNAs with a highly significant prognostic value in each cancer type. In total, we found 436 differentially expressed SE-lncRNAs and  $\sim$ 53% of these molecules were dysregulated in one cancer type, which may reflect their tissue-specific regulation in tumorigenesis (Table 2 and Figure 2A). In addition to these dysregulated SE-lncRNAs, we identified 2035 SE-lncRNAs with a highly significant prognostic value by using Cox's proportional hazard model (Table 2). Similar to the dysregulated genes,

most of the SE-lncRNAs with superior prognostic value showed the cancer type-specific pattern (Table 2 and Figure 2B).

To reveal the potential functions of SE-lncRNAs in cancers, it is important to identify the genes that are truly regulated by the SE-lncRNAs in each cancer type. As the close correlation between the expression of SE-lncRNAs and their regulated genes in cancers may reflect their regulatory relationships, we next evaluated their relationships by calculating their Pearson's correlation coefficients for each cancer type. We found that 3622 cis-acting regulatory relationships and 401 trans-acting regulatory relationships were significantly correlated, which may reveal the truly regulated genes of SE-lncRNAs in cancers (Table 2). For some lncRNAs for which the function is known, we found that the pathological functions of 91 SE-lncRNAs were annotated by the LncRNADisease and Lnc2Cancer databases.

#### Database introduction

To store and display the related data of SE-IncRNAs, a database SELER was built. The search results of SELER consisted of the following three sections: the putative SE-IncRNAs section, cancer-related SE-IncRNAs section and function known SE-IncRNA section. The putative SE-IncRNAs section offered basic information on SEs, SE-IncRNAs and their putative regulated genes (Figure 3A). The cancer-related SE-IncRNAs section provided their expression profiles, prognostic values and significantly correlated genes in different cancers (Figure 3B). The function-known SE-IncRNA section provided the experimentally validated functions of SE-IncRNAs in diseases. Apart from being shown in the database, all of the abovementioned





SE-lncRNAs with highly prognostic value



А									В																		
ENST00000546421.1_1	TMPO-AS1-201	chr12:98909072- 98909988(-)	SE_15101	chr12:9889	6418-98915734	CD4_Memory_I	Pri 1	Details	Expres	ision fold	l change	betweer	n tumoi	tissues	and nor	mal tissu	Fold cha	ncRNA g	tene: TN	IPO-AS	1)						
Super enhancer	Ensembl ID	Symbol	Gene coordinate	,	Distance	1	Regulated ger	ne type	0.47																		
SE_15101	ENST00000501499	RP11-181C3.1	chr12:98879322-	98897633(-)	18101		Gene TSS neighboring a SE		0.40																		
SE_15101	ENST00000549004	RP11-181C3.1	chr12:98880751-	98884864(-)	11554		Gene TSS neigh	boring a SE	0.35																		
SE_15101	ENST00000556029	тмро	chr12:98909290- 98944157(+)		12872	Gene TSS neighboring a SE		0.30																			
SE_15101	ENST00000548911	тмро	chr12:98909357- 98921791(+)		12939	Gene TSS neighboring a SE		0.25																			
SE_15101	ENST00000343315	тмро	chr12:98909396- 98942606(+)		12978		Gene TSS neigh	boring a SE	0.20																		
SE_15101	ENST0000266732	тмро	chr12:98909408- 98929412(+)		12990		Gene TSS neigh	iboring a SE	0.15																		
SE_15101	ENST00000393053	тмро	chr12:98909409- 98942735(+)		12991		Gene TSS neigh	iboring a SE	0.10																		
SE_15101	ENST00000546828	тмро	chr12:98909410- 98925557(+)		12992		Gene TSS neigh	boring a SE	0.05																		
SE_15101	ENST0000261210	тмро	chr12:98909630- 98931491(+)		13212		Gene TSS neigh	boring a SE		BLCA	BRCA	CESC	н	NSC	KICH	KIRC	Can	cer type	IC .	LUAD	LUSC	PRAD		STAD	THCA	UCI	EC.
SE_15101	ENST00000549938	тмро	chr12:98910030- 98926744(+)		13612		Gene TSS neigh	boring a SE	The X-au	kis means o	ifferent ci	ancer types	s and the	values of Y	-axis mea	in the fold	change (tu	mor/norm	al) of SE-I	ncRNAs. 1	his section	on shows th	e expre	ssion of 5	SE-IncRN4	A gene.	
SE_15101	ENST00000552831	тмро	chr12:98921307- 98931352(+)		5573		Gene TSS neigh	boring a SE	Progr	BRCA	CESC	n (SE-Inc	RNA g	KIRC	KIRP	) LIHC	LUAD	LUSC	PRAD	STAD	тно	A UCE	C RE	EAD L	LGG (	ov	GBM
SE_15101	ENST00000547214	тмро	chr12:98921663- 98926050(+)		5929		Gene TSS neigh	boring a SE	0	0	•	0	•	•	0	•	•	0	0	0	0	0	0	¢	<b>)</b> (	D	0
SE_15101	ENST00000556678	тмро	chr12:98921664- 98941479(+)		5930		Gene TSS neigh	aboring a SE	■ memory account on a minimum program or programmers and prove that an emerginal account of the memory acc																		
SE_15101	ENST00000548223	тмро	chr12:98937980- 98941642(+)		22246		Gene TSS neigh	boring a SE	Correlated gene (Pearson correlation, P-value<0.05, SE-IncRNA gene: TMPO-AS1)																		
SE_15101	ENST00000551987	тмро	chr12:98938417-		22683		Gene TSS neigh	boring a SE	Gene	BLCA	BRCA	CESC C	CAD	GBM H	NSC KI	CH KIR	C KIRP	LIHC	LUAD	LGG	LUSC	OV P	RAD	READ	STAD	THCA	UCEC
			78741565(+)						IMPO	0.301	0.007	0.077 0		0.504 0.	120 0	0.00	0.040	0.007	0.708	0.033	0.007	0.721 0	~~~	0.401	0.000	0.000	0.355

Figure 3. Sample output diagram for the result of the SE-IncRNA section. (A) Information about super enhancers, SE-IncRNAs and their associated genes. (B) Cancer-related information about SE-IncRNAs, including their expression profiles, prognostic information and significantly associated genes in cancers.

data could be downloaded from the download interface of SELER.

To identify cancer-related SE-lncRNAs and to explore their potential functions, users could apply filtering options to select significantly differentially expressed SE-lncRNAs with a high prognostic value. Moreover, the correlation coefficient between SE-lncRNAs and their regulated genes could help users to identify their truly regulated genes. For instance, we first chose the filtering options of DF&PV option to retain significantly differentially expressed cis-acting SE-lncRNAs with highly prognostic value. Then, we input TMPO-AS1-201 to search the database. We found that TMPO-AS1-201 was significantly differentially expressed in seven cancer types (Figure 3B). More importantly, TMPO-AS1-201 showed a high prognostic value in two of the seven cancer types, including liver hepatocellular carcinoma (LIHC) and lung squamous cell carcinoma (LUSC) (Figure 3B). To reveal the potential functions of TMPO-AS1-201 in these cancers, we investigated its significantly correlated genes in corresponding cancers and found that it was significantly correlated with thymopoietin (TMPO) in LIHC and LUSC (Figure 3B), which indicated that TMPO-AS1-201 may participate in tumorigenesis by regulating the gene expression of TMPO (Figure 3B).

# Discussion

Multiple experimental and computational data were integrated to identify cancer-related SE-lncRNAs and to explore their potential regulatory functions in tumorigenesis. In total, 2122 cancer-related SE-lncRNAs were identified, including 436 significantly differentially expressed lncR-NAs and 2035 with a high prognostic value. Moreover, 3935 significantly correlated relationships between SElncRNAs and their regulated genes were identified in 19 cancer types. Finally, the SELER database was built to provide users with a useful tool to investigate SE-lncRNA functions in cancers.

Compared to other lncRNA databases, the distinctive features of SELER are as follows: (i) SELER mainly focused on SE-lncRNAs, which regulated the gene expression by controlling SE activity. Our database systematically identified SE-lncRNAs and their putative regulated genes. (ii) SELER integrated large amounts of expression profile data sets and prognostic information derived from 19 cancer types to identify cancerassociated SE-lncRNAs. (iii) SELER comprehensively annotated SE-lncRNAs for which the functions are known. (iv) SELER provided a user-friendly interface for users to explore the potential roles of SE-lncRNAs in tumorigenesis.

In summary, SELER is a novel database that integrates large amounts of experimental and computational data to decode the regulated functions of SE-lncRNAs. Considerable information is offered to facilitate the investigation of SE-lncRNAs in tumorigenesis. SELER is expected to improve our comprehensive understanding of the important and novel roles of SE-lncRNAs in the regulation of gene expression and in pathological processes.

# Supplementary data

Supplementary data are available at Database Online.

# Funding

Science and Technology Program of Guangdong [2015B020233009, 2015A030401040]; China Postdoctoral Science Foundation funded project [2016M602486]; National Science Foundation for Young Scientists of China [81802435].

## References

- Pott,S. and Lieb,J.D. (2015) What are super-enhancers? Nat. Genet., 47, 8–12.
- Khan,A. and Zhang,X. (2016) dbSUPER: a database of superenhancers in mouse and human genome. *Nucleic Acids Res.*, 44, D164–D171.
- 3. Mousavi,K., Zare,H., Dell'orso,S. *et al.* (2013) eRNAs promote transcription by establishing chromatin accessibility at defined genomic loci. *Mol. Cell*, **51**, 606–617.
- Ong,C.T. and Corces,V.G. (2011) Enhancer function: new insights into the regulation of tissue-specific gene expression. *Nat. Rev. Genet.*, 12, 283–293.
- Mack,S.C., Pajtler,K.W., Chavez,L. *et al.* (2018) Therapeutic targeting of ependymoma as informed by oncogenic enhancer profiling. *Nature*, 553, 101–105.
- Katerndahl,C.D.S., Heltemes-Harris,L.M., Willette,M.J.L. *et al.* (2017) Antagonism of B cell enhancer networks by STAT5 drives leukemia and poor patient survival. *Nat. Immunol.*, 18, 694–704.
- Iyer,S., Modali,S.D. and Agarwal,S.K. (2017) Long noncoding RNA MEG3 is an epigenetic determinant of oncogenic signaling in functional pancreatic neuroendocrine tumor cells. *Mol. Cell. Biol.*, 37, e00278–17.
- Berretta, J. and Morillon, A. (2009) Pervasive transcription constitutes a new level of eukaryotic genome regulation. *EMBO Rep.*, 10, 973–982.
- Hangauer,M.J., Vaughn,I.W. and McManus,M.T. (2013) Pervasive transcription of the human genome produces thousands of previously unidentified long intergenic noncoding RNAs. *PLoS Genet.*, 9, e1003569.
- Ning,S., Zhang,J., Wang,P. et al. (2016) Lnc2Cancer: a manually curated database of experimentally supported lncRNAs associated with various human cancers. Nucleic Acids Res., 44, D980–D985.
- 11. Chen,G., Wang,Z., Wang,D. *et al.* (2013) LncRNADisease: a database for long-non-coding RNA-associated diseases. *Nucleic Acids Res.*, **41**, D983–D986.
- Schaukowitch,K., Joo,J.Y., Liu,X. *et al.* (2014) Enhancer RNA facilitates NELF release from immediate early genes. *Mol. Cell*, 56, 29–42.
- Soibam,B. (2017) Super-IncRNAs: identification of IncRNAs that target super-enhancers via RNA:DNA:DNA triplex formation. RNA, 23, 1729–1742.
- Anderson,K.M., Anderson,D.M., McAnally,J.R. *et al.* (2016) Transcription of the non-coding RNA upperhand controls Hand2 expression and heart development. *Nature*, 539, 433–436.
- Ounzain,S., Micheletti,R., Arnan,C. *et al.* (2015) CARMEN, a human super enhancer-associated long noncoding RNA controlling cardiac specification, differentiation and homeostasis. *J. Mol. Cell. Cardiol.*, 89, 98–112.
- Xiang, J.F., Yin, Q.F., Chen, T. *et al.* (2014) Human colorectal cancer-specific CCAT1-L lncRNA regulates long-range chromatin interactions at the MYC locus. *Cell Res.*, 24, 513–531.
- Harrow, J., Frankish, A., Gonzalez, J.M. *et al.* (2012) GENCODE: the reference human genome annotation for The ENCODE Project. *Genome Res.*, 22, 1760–1774.

- Zhen, C., Pan, X., Yang, Y. *et al.* (2018) The IncLocator: a subcellular localization predictor for long non-coding RNAs based on a stacked ensemble classifier. *Bioinformatics*, 34, 2185–2194.
- 19. Buske,F.A., Bauer,D.C., Mattick,J.S. *et al.* (2012) Triplexator: detecting nucleic acid triple helices in genomic and transcriptomic data. *Genome Res.*, 22, 1372–1381.
- 20. Chepelev,I., Wei,G., Wangsa,D. *et al.* (2012) Characterization of genome-wide enhancer-promoter interactions reveals coexpression of interacting genes and modes of higher order chromatin organization. *Cell Res.*, **22**, 490–503.
- 21. Mi,H., Muruganujan,A. and Thomas,P.D. (2013) PANTHER in 2013: modeling the evolution of gene function, and other gene attributes, in the context of phylogenetic trees. *Nucleic Acids Res.*, **41**, D377–D386.
- 22. Li,J., Han,L., Roebuck,P. *et al.* (2015) TANRIC: an interactive open platform to explore the function of lncRNAs in cancer. *Cancer Res.*, **75**, 3728–3737.